



Canine Babesiosis: An Overview

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Abstract

Canine Babesiosis is one of the most common, globally existent, fast spreading tick-borne diseases of haemoprotozoan origin caused by different species of Babesia. The clinical signs appear as a result of hemolysis due to presence of the organism within the erythrocytes. But some species of Babesia may also trigger immune mediated component to anaemia along with severe inflammatory reaction resulting to morbidity and mortality in animals. The various species of Babesia are *B. canis*, *B. vogeli*, *B. microti*, *B. rossii* and *B. gibsoni*. Canine Babesiosis occurs mainly due to two forms of Babesia namely Large form and small form. Within large form *Babesia canis* is reported and in small form *B. gibsoni* is the main causative agent. In this article the taxonomy, geographical distribution, transmission, clinical signs, diagnosis, treatment and prevention on canine babesiosis is discussed.

Received: Jul 20, 2021

Accepted: sep 08, 2021

Published Online: Sep 10, 2021

Journal: Journal of Veterinary Medicine and Animal Sciences

Publisher: MedDocs Publishers LLC

Online edition: <http://meddocsonline.org/>

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Keywords: Babesia; Ticks; Treatment.

Introduction

In the year 1888, Dr. Victor Babes, a Romanian physician, was the first to observe microorganisms in the erythrocytes of sheep and cattle showing the symptoms of haemoglobinuria [1]. Later these micro-organisms were named *Babesia ovis* and *Babesia bovis* respectively. In 1895, not long after the observation in ruminants, the same type of observations ie. *Babesia* spp. for the first time in dogs came into light in Italy [2]. Currently this protozoan disease is prevalent in various parts of the world.

Canine babesiosis was formerly known as Canine piroplasmosis. The *Babesia* spp. is mainly transmitted by hard ticks. An important phase of life cycle of *Babesia* spp. ie. sexual conjugation and sporogony takes place within the intestinal lumen fol-

lowed by haemocoel of the ticks. Ultimately a blood meal transmits the sporozoites from the tick's salivary gland to a new host wherein the protozoan life cycle is completed by asexual replication or merogony within the erythrocytes where the parasites remain as merozoites.

Taxonomy and morphology

The *Babesia* genus belongs to the order *Piroplasmida* in the phylum *Apicomplexa* and can be seen as a non-pigmented pear or signet ring shaped organisms in the mammalian erythrocytes. The large forms of *Babesia* (2.5-5.0 µm) consists three species viz., *B. canis*, *B.vogeli* and *B. rossii* and the small forms (1.0–2.5 µm) comprising of the. *B. gibsoni*, *B. conradae* and *B. microti* like piroplasms [3] (Figure 1 & 2).

Cite this article: Halder B, Gupta AR. Canine Babesiosis: An Overview. J Vet Med Animal Sci. 2021; 4(2): 1081.



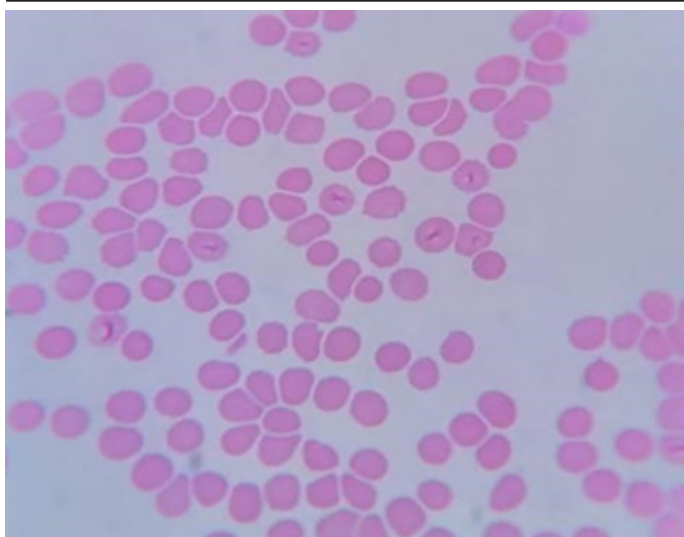


Figure 1: *Babesia canis* (Large form).

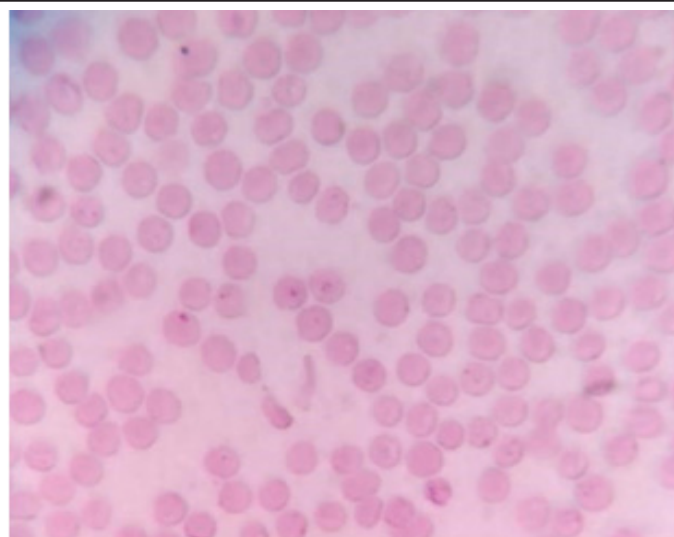


Figure 2: *Babesia gibsoni* (Small form).

Geographical distribution and transmission

Canine Babesiosis is clinically significant haemoprotozoan disease of dogs which is distributed world-wide including India. The disease appears during the whole year period with frequent outbreaks in the spring and autumn. The disease is transmitted through tick bites. Trans-stadial and trans-ovarial both transmission may occur and the ticks remain infective for several generation. Babesiosis can also be transmitted by blood transfusion. Recently proven that trans-placental transmission occur from dam to offspring [4]. *B. gibsoni* can also be transmitted by dog bites [5].

The species of *Babesia* transmission by vectors and their geographical distribution is given in the table below [6,7].

Species	Vector	Geographical Distribution
<i>Babesia canis</i>	<i>Dermacentor reticulatus</i>	Europe
<i>B. vogeli</i>	<i>Rhipicephalus sanguineus</i>	Tropical and subtropical region
<i>B. rossi</i>	<i>Haemophysalis elliptica</i>	South Africa
<i>B. gibsoni</i>	<i>Haemophysalis sp.</i>	Asia, North America, Northern and Eastern Africa and Europe
<i>B. microti</i>	<i>Ixodes hexagonus Ixodes canisuga</i>	France, Croatia, Italy, Portugal, Serbia, Spain and Sweden

Clinical signs

In general clinical signs of canine babesiosis are dependent on species, breed, age, immune status of animal and concurrent disease [8]. The disease can be classified as complicated and uncomplicated forms [9]. Uncomplicated forms can be categorized by haemolytic anaemia accompanied with inappetance to complete anorexia, increased pulse rate, heart rate, palor mucous membrane, pyrexia and in some case epistaxis is detectable. Complicated form characterized by Systemic Inflammatory Response Syndrome (SIRS) and Multiple Organ Dysfunction Syndrome (MODS), both of which are cytokine mediated phenomenon [10,11]. Both uncomplicated and complicated Babesiosis appear to be the result of host inflammatory responses [12,13]. The immunological response plays the most important role in pathogenesis of canine babesiosis. This parasite initiates the mechanism of antibody mediated cytotoxic destruction of circulating erythrocytes. Autoantibodies are directed against com-

ponents of the membranes of infected and uninfected erythrocytes which causes intravascular and extravascular haemolysis.

Diagnosis

Microscopical examination

Most commonly used diagnostic method is direct microscopical examination of thin blood smear as it is cost effective, feasible and conclusive method. For microscopical examination, blood is collected from ear vein and a thin smear is made. It is then heat fixed and Giemsa staining is done following the standard protocol. In positive cases pear-shaped piroplasms are detected and the differentiation of species is done on the basis of size and appearance inside the erythrocytes [14].

Haematological findings

Haematological changes also examined in canine babesiosis. Whole blood is collected into the EDTA vial for blood cell parameters detection. There is mild to moderately regenerative normocytic and normochromic anaemia because of haemolysis. Haematological examination revealed elevated haemoglobin concentration, higher lymphocytes, monocytes and eosinophil count. Neutrophil counts are usually normal to decreased and left shifts are seen. Thrombocytopenia is a hallmark sign of this disease [15] and is usually severe in the acute phase of infection [16].

Biochemical profile

The serum sample is analysed for examination of biochemical profile. Total protein, albumin, Albumin/Globulin ratio, glucose level are reduced [17]. Liver enzymes such as ALP, ALT, AST and bilirubin level are also elevated with marked icterus due to cellular damage to the hepatic cells [18]. Urea and creatinine levels are also increased. Increased level of ALP may be occurs due to damage or abnormal function of the biliary system. This might be due to liver or kidney involvement. Serum potassium is reduced, especially in the icteric cases. Azotemia is present in dehydrated cases and in those with acute renal failure [19]. The changes may be occurs due to hepatopathy and immune haemolytic anaemia which is caused by the organism.

Serological test

For confirmatory diagnosis of canine babesiosis serological test is done. Active infection can be confirmed by increasing

antibody titers detection. The serological method, indirect fluorescent antibody test (IFAT) and enzyme linked immuosorbent assay (ELISA) are highly sensitive for *B. gibsoni* but moderately specific to *B. canis* due to antigenic cross- reaction [20].

Molecular techniques

Polymerase Chain Reaction (PCR) offers a practical and non-invasive means to detect and differentiate infections with various *Babesia* spp. and also provides a sensitive tool for assessing treatment outcomes. Different molecular techniques are used for the identification and differentiation of the various species of *Babesia* viz., semi-nested PCR [21], reverse line blotting [22,23], and PCR- restriction fragment length polymorphism analysis [24]. Moreover, sequencing of 18S rRNA and Internal Transcribed Spacer-1 (ITS-1) have also been used for molecular phylogeny studies of this parasite [25].

Treatment

The treatment of babesiosis involves the removal of parasite from the body, correction of anaemia along with supportive treatment. Diminazene aceturate, imidocarb dipropionate and trypan blue are effective against large form of babesiosis. Diminazene aceturate @3.5 mg/kg body weight is administered subcutaneously or intra-muscularly but it has some toxic adverse effects with severe neurological signs [26]. Imidocarb dipropionate @6.6 mg/kg body weight 14 days apart is also widely included in treatment protocol and is injected intra-muscularly [26,27]. Administration of Imidocarb may cause pain at the injection site and reveals cholinergic signs like salivation, vomiting, diarrhoea, lacrimation, nasal drip etc. Cholinergic side effects may be controlled by administration of atropine sulfate @0.04 mg/kg body wt. subcutaneously. Trypan blue @10 mg/kg wt. one dose administered intravenously followed by diminazene or imidocarb 1 week later was earlier used and is still being used in some parts of the world [6].

In the case of *B. gibsoni*, Diaminazene aceturate, satisfactory results are not obtained as the drug is unable to remove the parasite completely. It is treated with combination of three drugs consisting of Doxycycline @5 mg/kg body wt. orally twice daily, Clindamycin @25 mg/kg body wt. orally twice daily and Metronidazole @15 mg/kg body wt. orally twice daily or Doxycycline @ 7-10 mg/kg body wt. orally twice daily, Enrofloxacin @ 2-2.5 mg/kg body wt. orally twice daily and Metronidazole @ 5-15 mg/kg orally twice daily [28]. Amphotericin B has been shown activity against *B. gibsoni* but it caused oxidative red blood cell damage *in-vitro* and kidney damage *in-vivo* [28].

Buparvaquone @5 mg/kg body wt. intramuscularly repeated after 48 hours in combination with Azithromycin @10 mg/kg body wt. once daily orally for 10 days [27,29]. But the side effect of this treatment protocol was found to be allergic reaction and itching which can be controlled by Dexamethasone @0.5 mg/kg body wt. intramuscularly along with Chlorpheniramine maleate @0.5 mg/kg intramuscularly. The other combination is Atovaquone @13.3 mg/kg body wt. thrice daily orally for 10 days in combination with Azithromycin @10 mg/kg body wt. once daily orally for 10 days [30].

Prevention

As it is a tick-borne disease so tick control is the most important factor. Regular examination of dogs to remove the tick is important for protection of animals from babesiosis. Various types of topical products e.g. tick control spray, shampoo, pow-

der are recently available in market to control the ticks. Ivermectin @0.2 mg/kg body weight subcutaneously at 1 week interval is effective to get rid of ticks from the body of host. The use of amitraz impregnated collars give satisfactory result to control the *Babesiosis* infection in endemic areas. Awareness of the owners regarding tick control helps in eradication of the disease as early as possible.

In Europe a vaccine is available against *B. canis* having 70-100 % of efficacy. Recently a bivalent vaccine is derived from soluble parasite antigens from *B. canis* and *B. rossi* which helps to reduce the duration and severity of infection [28]. Although the vaccination doesn't prevent the infection but it blocks the initiation of pathogenic processes involved in the pathogenesis of the diseases.

References

1. Babes V. Sur l'haemoglobinurie bacterienne duboef. C R Hebd Seances Acad Sci. 1888; 107: 692-694.
2. Roncalli AR. The history of Italian parasitology. Vet Parasitol. 2001; 98: 3-30.
3. Irwin PJ. Canine babesiosis: from molecular taxonomy to control. Parasit Vectors. 2009; 2: 1-9.
4. Fukumoto S., Suzuki W., Igarashi I., Xuan X. Fatal experimental transplacental *Babesia gibsoni* infections in dogs. Int J Parasitol. 2005; 35: 1031-1035.
5. Birkenheuer AJ, Correa MT, Levy MG, Breitschwerdt EB. Geographic distribution of Babesiosis among dogs in the United States and association with dog bites: 150 case (2000-2003). J Am Vet Med Assoc. 2005; 227: 942-947.
6. Schoeman JP. Canine babesiosis. Onderstepoort J Vet Res. 2006; 76: 59-66.
7. Gallego L, Sainz A, Roura X, Estrapada-Pena M. A review of canine babesiosis: the European perspective. Parasit Vectors. 2016; 9: 336.
8. Asokkumar M, Selvaraju G, Naveen Kumar V. Canine babesiosis in a Doberman Dog- Successive Therapeutic Management. Shanlax Int J Vet Sci. 2017; 4: 23-25.
9. Yogespriya S, Pillai UN, Ajithkumar S. Successful Management of Canine Babesiosis- A Case Report. Shanlax Int J Vet Sci. 2014; 3: 33-34.
10. Jacobson L, Clark SIA. The pathophysiology of canine babesiosis: new approaches to an old puzzle. J S Afr Vet Assoc. 1994; 65: 134-145.
11. Welzl C, Leisewitz AL, Jacobson LS, Vaughan Scott T, Myburgh E. Systemic inflammatory response syndrome and multiple organ damage/dysfunction in complicated canine babesiosis. J S Afr Vet Assoc. 2001; 72: 158-162.
12. Motijatko V, Mrljak V, Kis I, Kucer N, Forseck J, et al. Evidence of acute phase response in dogs naturally infected with *Babesia canis*. Vet Parasitol. 2007; 144: 242-250.
13. Schettters TP, Kleuskens JA, Van De Crommert J, De Leeuw PWJ, Finzio AL, et al. Systemic inflammatory responses in dogs experimentally infected with *Babesia canis*; a haematological study. Vet Parasitol. 2009; 162: 7-15.
14. Laha R, Bhattacharjee K, Sarmah PC, Das M, Goswami A, et al. *Babesia* infection in naturally exposed pet dogs from a north-eastern state (Assam) of India: detection by microscopy and polymerase chain reaction. J parasitic dis. 2014; 38: 389-393.

15. Salem NY, Farag HS. Clinical, hematologic, and molecular findings in naturally occurring *Babesia canis vogeli* in Egyptian dogs. *Vet Med Int.* 2014; 270345.
16. Furlanello T, Fiorio F, Caldin M, Lubas G, Solano-Gallego L. Clinicopathological findings in naturally occurring cases of babesiosis caused by large form *Babesia* from dogs of Northern Italy. *Vet. Parasitol.* 2005; 134: 77-85.
17. Reddy BS, Vivajothi S, Reddy LSSV, Raju KGS. Clinical and laboratory findings of *Babesia* infection in dogs. *J parasitic dis.* 2016; 40: 268-272.
18. Praveen K, Abhishek K. Haemato-biochemical changes in dogs infected with Babesiosis. *Int J Chem Studies.* 2018; SP4: 25-28.
19. Lobetti RG, Jacobson LS. Renal involvement in dogs with babesiosis. *J S Afr vet Ass.* 2001; 72: 23-28.
20. Amritpal S, Harkirat S, Singh NK, Singh ND, Rath SS. Canine babesiosis in Northwestern India: Molecular detection and Assessment of Risk Factors. *J Biomed Biotech.* 2014; 1: 741785.
21. Birkenheuer AJ, Levy MG, Breitschwerdt EB. Development and evaluation of a seminested PCR for detection and differentiation of *Babesia gibsoni* (Asian genotype) and *B. canis* DNA in canine blood samples. *J Clin Microbiol.* 2003; 41: 4172-4177.
22. Matjila PT, Penzhorn BL, Bekker CP, Nijhof AM, Jongejan F. Confirmation of occurrence of *Babesia canis vogeli* in domestic dogs in South Africa. *Vet Parasitol.* 2004; 122: 119-125.
23. Yisaschar-Mekuzas Y, Jaffe CL, Pastor J, Cardoso L, Baneth G. Identification of *Babesia* species infecting dogs using reverse line blot hybridization for six canine piroplasms, and evaluation of co-infection by other vector-borne pathogens. *Vet Parasitol.* 2013; 191: 367-373.
24. Jefferies R, Ryan UM, Irwin PJ. PCR-RFLP for the detection and differentiation of the canine piroplasm species and its use with filter paper-based technologies. *Vet Parasitol.* 2007; 144: 20-27.
25. Mandal M, Banerjee PS, Garg R, Ram H, Kundu K, et al. Genetic characterization and phylogenetic relationships based on 18S rRNA and ITS1 region of small form of canine *Babesia* spp. from India. *Infect Genet Evol.* 2014; 27: 325-331.
26. Plumb DC. *Plumb's Veterinary Drug Handbook*, 8th edition. Wille-Blackwell, Ames. 2015; 1296.
27. Baneth G. Antiprotozoal treatment of canine babesiosis. *Vet Parasitol.* 2018; 254: 58-63.
28. Köster LS, Lobetti RG, Kelly P. Canine babesiosis: a perspective on clinical complications, biomarkers, and treatment. *Vet med (Auckland, N.Z.).* 2015; 6: 119-128.
29. Checa R, Montoya A, Ortega N, Gonzalez-Fraga JL, Bartolome A, et al. Efficacy, Safety and tolerance of imidocarb dipropionate versus Atovaquone or buparvaquone plus Azithromycin used to treat sick dogs naturally infected with the *Babesia microti*-like piroplasm. *Parasit Vectors.* 2017; 10: 145.
30. Birkenheuer AJ, Levy MG, Breitschwerdt EB. Efficacy of combined Atovaquone and Azithromycin for therapy of chronic *Babesia gibsoni* (Asian genotype) infections in dogs. *J Vet Int Med.* 2004; 18: 494-498.