

**Annals of Pharmaceutical Sciences** 

**Open Access | Review Article** 

# A Review on Phytochemical & Pharmacological Properties of Bidens Bipinnata

## Muhammad Murtaza\*; Mohd Asif

Glocal University Pharmacy College, Glocal University Saharanpur, India.

## \*Corresponding Author(s): Muhammad Murtaza

Glocal University Pharmacy College, Glocal University Saharanpur, India. Email: murtazachoudhary5@mail.com

Received: Dec 14, 2025 Accepted: Jan 29, 2025 Published Online: Feb 05, 2025 Journal: Annals of Pharmaceutical Sciences Publisher: MedDocs Publishers LLC Online edition: http://meddocsonline.org/ Copyright: © Murtaza M (2025). This Article is distributed under the terms of Creative Commons Attribution 4.0 International License

**Keywords:** Bidens Bipinnata; Phytochemical constituents; Traditional uses; Isolated compound; Pharmacological activity.

#### Abstract

Bidens bipinnata Linn, also known as B. bipinnata, is a member of the Asteraceae family that has been utilized for various medicinal purposes for a very long time in a variety of different civilizations. The purpose of this research is to gather the most recent information that is currently available on the phytochemical components and pharmacological activities of *B. bipinnata*. It is believed that the range of bioactive components that it possesses, such as flavonoids, terpenoids, and phenolics, are responsible for the medicinal properties that it possesses. Studies in the field of pharmacology have demonstrated that B. bipinnata possesses many pharmacological activity, such as anticancer, antibacterial, anti-inflammatory, and antioxidant characteristics. In particular, the total flavonoids found in B. bipinnata have been shown to have significant neuroprotective qualities in brain damage models, which makes it a potential candidate for more pharmacological investigation. The present study provides preliminary data as well as recommendations for more research in the fields of science and clinical medicine pertaining to this plant.

#### Introduction

Plants have long been integral in treating diverse ailments globally. The demand for medicinal plants is rising in developed and developing nations alike, owing to their minimal adverse effects. Herbal medicine constitutes a significant component of both traditional and contemporary medical systems [1]. Medicinal plants are still important in healthcare systems in many different parts of the world, where 80 percent of people rely mostly on traditional medicine to meet their medical requirements. Over four billion people, or 80% of the world's population, live in poor countries and depend mostly on plant-derived products for their healthcare needs [2]. Moreover, a considerable portion of contemporary medications utilized in modern medicine for treating illnesses, including aspirin, codeine, and quinine, have roots in traditional herbal remedies and medicinal plants. Consequently, medicinal plants serve as crucial reservoirs of highly effective medicines for treating illnesses and promoting longterm human health [3].

The presence of many bioactive chemicals, including lignans, tannins, terpenoids, polyphenols, alkaloids, and other secondary metabolites, is what gives medicinal plants their therapeutic qualities [4].

Bidens bipinnata (B. bipinnata), a member of the Asteraceae family, holds significant importance due to its numerous ethnomedicinal and nutritional properties. Traditionally, B. bipinnata is extensively utilized for treating a range of conditions, such as asthma, laryngeal and bronchial diseases, as well as for its styptic and vermifuge qualities, effective in addressing conditions like sore throat and conjunctivitis. Additionally, it is applied topically for wound healing, exhibiting bactericidal properties and demonstrating antimalarial activity, thereby aiding in alleviating symptoms such as fever, cough, and asthma [5].

Bidens pilosa and B. bipinnata are two of the species of the species group Bidens that have been the subject of much scientific research in recent decades on their possible medical



**Cite this article:** Murtaza M, Asif M. A Review on Phytochemical & Pharmacological Properties of Bidens Bipinnata. Ann Pharm Sci. 2025; 1(1): 1001. benefits. Numerous beneficial chemicals have been identified as a result of these research efforts, including  $\beta$  caryophyllene,  $\beta$ -carotene, sesquiterpenes, and germacrene-D [6].

Investigating *B. bipinnata's* potential therapeutic benefits requires tying in-depth scientific study with the plant's traditional medical usage. This review explores contemporary research on the phytochemistry, pharmacology, toxicity, traditional applications, and botany of the plant. The combined information demonstrates the possible benefits of *B. bipinnata* and its constituent parts and suggests directions for further study to produce strong medicinal molecules.

#### Botany

B. bipinnata is an annual plant predominantly found in the temperate biome. It serves various purposes such as animal fodder, a source of poison, medicinal applications, and as food for invertebrates. Additionally, it holds environmental significance and is utilized as human food. According to taxonomic classification Table 1 shows that it belongs to the Bidens genus (Asteraceae) [7]. There are thought to be 230-240 species in this genus globally [8]. B. bipinnata thrives in wetland, tropical islands, and various habitats including river bluffs, pine-oak forests, campsites, roadsides, and abandoned farms. It favors shady areas and wet sandy soils like sandy loam, red sandy clay, and loamy sand. In North Florida flatwoods forests, it remains unaffected by soil disturbance caused by clearcutting and chopping [9]. B. bipinnata, commonly known as Spanish Needles, is an annual herb that grows to be approximately 2 to 5 feet tall, almost glabrous in texture. Stem: 4-angled, frequently branching, roughly straight. The majority of the leaves are opposite and are long petiolate, with up to four pairs of strongly lobed pinnae and scanty hair on the veins underneath. The last segments might be whole or dentate, cuneate at the base, and ovate to rhombic-lanceolate. The centre of the flower head has tubular disc blooms, encircled by strap-shaped ray flowers that are either yellow or white in colour. During the flowering stage, the single, upright capitula measure 6-8.5 x 5-6.5 mm, and the peduncles vary in length from 2 to 10 cm. The outermost involucral bracts are herbaceous, linear-lanceolate to rectangular, and shorter than the inner ones. Shorter than achenes, receptacular scales have black longitudinal lines and are scarious and linear. The flowers are roughly the same length as the involucre and are made up of disc florets with yellowish corollas and 0-4 ligulate florets that are 2-3 mm long (Figure 1) [10]. Achenes are brown-blackish, ranging from 8-10 mm to 10-18 mm in length, linear in shape, with a pappus of (2) 3-4 bristles measuring 2-4 mm long. The plant flowers from August to October, with fruiting occurring from September to October. Commonly known as Spanish Needles, it is naturalized worldwide, probably native to the American continents [11,12].

Kingdom:	Plantae
Clade:	Tracheophytes
Clade:	Angiosperms
Clade:	Eudicots
Clade:	Asterids
Order:	Asterales
Family:	Asteraceae
Genus:	Bidens
Species:	B. bipinnata



Figure 1: Bidens Bipinnata (A): flower of B. Bipinnata. (B): Needle-like seeds, hence the name "Spanish Needle.

#### **Geographical distribution**

The geographical distribution of *B. bipinnata* spans across North and South America. It is native to Mexico, Central America, United States, Canada, South America. Common in Eastern and central regions of the United States Extends into Canada, Brazil, Argentina, Colombia, Venezuela, and other countries in South America (Figure 2).

B. bipinnata thrives across Florida, Georgia, Maryland, Kentucky, Louisiana, North America, Mississippi, North Carolina, Eastern Canada, South Carolina, Tennessee, and Virginia. It extends its reach to various regions, including Cyprus in Western Asia, the United Kingdom in Northern Europe, and Belgium, Germany, and Hungary in Middle Europe, with further presence noted in Estonia within Eastern Europe, and Montenegro and Romania in Southeastern Europe, and Spain in Southwestern Europe. Naturalized populations have established in diverse locations across Africa, Ghana, Guinea, Nigeria, Sierra Leone, Togo, Malawi, Zimbabwe, Botswana, Lesotho, Eswatini, and Madagascar in the Western Indian Ocean. Additionally, it has naturalized in regions such as Yemen on the Arabian Peninsula, the Caucasus region including Ciscaucasia within the Russian Federation and Georgia, China, Korea, and Taiwan in Eastern Asia, and Bhutan, India, and Nepal within the Indian Subcontinent. Its reach extends into Southeast Asia, including Cambodia, Laos, Thailand, and Vietnam in Indo-China, and the Philippines in Malesia. Furthermore, it has been recorded in Australia, specifically in the state of Australia, and across Europe, including Switzerland in Middle Europe, and Bulgaria, Croatia, Italy (including Sicily), and Slovenia in Southeastern Europe, with France in Southwestern Europe. In South America, it has been observed in Venezuela in Northern South America and Brazil, particularly in Rio Grande do Sul and Santa Catarina [13].



Figure 2: (Distribution of B. bipinnata) [14].

## **Ethnobotanical importance**

Table

*B. bipinnata* (Asteraceae) is traditionally utilized in China as an anti-inflammatory, antipyretic, and anti-rheumatic drugs, and it possesses a broad range of pharmacological applications [15]. As summarized in Table 1, *B. bipinnata* has been found to be beneficial in treating various diseases, including stomach ache, diarrhoea, dysentery, vaginitis, candidiasis, skin infections, and many others.

Diseases	References
Stomach Ache	[16]
Diarrhoea	[17]
Dysentery	[17]
Vaginitis	[18]
Candidiasis	[19]
Skin Infections	[17]

#### Table 2: Phytochemical Composition of B. Bipinnata.

## Phytochemical Composition of B. bipinnata

B. bipinnata exhibits a rich phytochemical profile, contributing to its diverse pharmacological applications. The analysis of its vegetative and reproductive parts reveals the presence of several bioactive compounds in significant quantities. The vegetative part contains 930±4.714 mg/100 g of tannins, which are known for their antioxidant properties. Alkaloids, which have therapeutic potentials such as analgesic and anti-inflammatory effects, are present at a concentration of 0.231%. Flavonoids, recognized for their role in reducing oxidative stress, are found at 5.09%. Saponins, which possess immunomodulatory and anti-cancer properties, are present at 7.09%. Oxalate content is measured at 2.97%, while cyanogenic glycosides, which can have both beneficial and toxic effects depending on the dose, are present at 520±0.275 mg/100 g. Phenols, essential for their antioxidant and anti-inflammatory activities, are found at 4.04±1.6750 mg/g, and lipids, which are vital for various physiological functions, are present at 6.4%.

In the reproductive part, tannins are slightly lower at  $865\pm0.9428 \text{ mg}/100 \text{ g}$ , while alkaloids are present at 0.201%. Flavonoid content is higher at 8.2%, and saponins are also more abundant at 9.8%. Oxalate content decreases to 1.76%, and cyanogenic glycosides are measured at  $465\pm1.3568 \text{ mg}/100 \text{ g}$ . Phenols increase to  $5.98\pm0.7979 \text{ mg/g}$ , while lipid content is slightly lower at 5.76% (Table 2) [20]. This phytochemical diversity underpins the medicinal value of *B. bipinnata* and supports its traditional use in treating various ailments [20].

Plant	Tannin (mg/100 g)	Alkaloid (%)	Flavonoid (%)	Saponins (%)	Oxalate (%)	Cyanogenic Glycoside (mg/100 g)	Phenols (mg/g)	Lipid (%)
Vegetative part	930±4.714	0.231	5.09	7.09	2.97	520±0.275	4.04±1.6750	6.4
Reproductive part	865±0.9428	0.201	8.2	9.8	1.76	465±1.3568	5.98±0.7979	5.76

#### **Compound isolation**

The mass spectrum of linoleic acid isolated from B. bipinnata shows a peak at m/z 280.4, which is consistent with the formula C18H32O2. This polyunsaturated omega-6 fatty acid is mainly insoluble in water, however it is soluble in organic solvents and colourless or white. Dehydroabietic acid is another active molecule with the chemical formula C20H28O2 with a molecular ion peak at m/z 300. It is a pyran-2,4-dione in which positions 3 and 6 have been replaced, respectively, with acetyl and methyl groups. Dehydroabietic acid belongs to the category of pyrone derivatives, which are heterocyclic substances made up of an oxygen atom, a ketone, and an unsaturated six-membered ring [20]. Linoleic acid is particularly significant for illness prevention, and it can prevent a number of allergic, depressive symptoms, cardiac, and some neurological conditions [21]. It has also been connected to improved membrane function and decreased blood cholesterol [22]. Linoleic acid has the most potent bactericidal effect against H. pylori, completely eliminating the germs via increasing the permeability of its outer membrane. This was validated by measuring released ATP, which indicated increased plasma membrane permeability and caused bacterial cell death. Transmission Electron Microscopy (TEM) and Scanning Electron Microscopy (SEM) revealed that linoleic acid altered the bacterial membrane's structure in under five minutes, affecting its integrity and resulting in cytoplasmic leakage [23]. One common chemical defence agent is dehydroabietic acid. It functions biologically in a variety of ways, including cytotoxic, antitumor, antiulcer, antiplasmodial, cardiovascular, antibacterial, antioxidant, and anti-inflammatory properties [24]. During the study on B. bipinnata, thirty-eight compounds were found and their structures were reported. These compounds included nine ceramides, thirteen flavonoids, five phenylpropanoids, four aliphatics, one pyrimidine, four steroids, one triterpenoid, and one polyacetylene. First, nine known compounds (4-9, 25, 26, 38) and five new compounds (1-3, 10-11) were taken out of the Bidens genus. Furthermore, new descriptions are provided for B. bipinnata Linn. compounds 12, 13, 16-21, and 30-34. The results are summarized in Table 3 [25]. A new flavanone glucoside named bidenoside F and a chalcone glucoside called bidenoside G have been isolated and structurally elucidated from B. Bipinnata [27]. The polyacetylenes that were isolated from B. bipinnata exhibited a range of characteristics and chemical makeups. Compound 1-also known as (2S)-(5E,11E)tridecadiene-7,9-diyne-1,2,13-triol-displayed distinct spectrum characteristics that provided insight into its structure and composition. Compound 2, also known as (6E, 12E)-3-oxo-tetradecadiene-8,10-diyne-14-hydroxyl-1-O-b-D-glucopyranoside,

had structural characteristics with the addition of a glucopyranose group. Both of these compounds and their recognized variants were subjected to in vitro testing to determine their cytotoxic and anti-inflammatory activities. Compounds 1, 5, and 6 had strong anti-inflammatory properties by reducing IL-1 and TNF-a levels, however none of them shown cytotoxicity against tested cancer cell lines [28]. Extract from *B. bipinnata* yielded a variety of chemicals with different structures. Compound 1 was recognized as a novel chlorinated flavonoid that is seldom seen in plants, namely 3,6,8-trichloro-5,7,3',4'-tetrahydroxyflavone. The unique chemical formula of compound 2, a phenylpropanoid glycoside, was C28H28O12. Compound 3, which exhibits unique structural properties, was identified as 8,3',4'-trihydroxyflavone-7-O- $\beta$ -D-glucopyranoside. Furthermore, the compounds isolated from *B. bipinnata*'s ethyl acetate

Table 3: Phytoconstituents of B. Bipinnata.

fraction were tested for their capacity to impede  $\alpha$ -amylase activity. Compound 6, isookanin, in particular, showed notable inhibitory action, with an IC50 value of 0.447 mg/ml [29]. The investigation into *B. bipinnata* effectiveness against hyperlipidemia involved analyzing its active components' tissue distribution and conducting molecular docking research. Compound isolation unveiled key molecules such as gallic acid, protocatechuic acid, rutin, hyperoside, Bipinnate Polyacetylenicloside (BPC), luteolin, and quercetin, recognized for their potential in treating hyperlipidemia. These compounds displayed significant distribution in liver tissue and exhibited binding affinity with multiple target proteins, suggesting their therapeutic promise against hyperlipidemia. Furthermore, the isolated polyacetylenes demonstrated noteworthy anti-inflammatory properties in vitro, underscoring their broader medicinal potential [30].

S.N.	Name	IUPAC Name	Class of Compound	Structure	References
1	-	(2S,3S,4R,8E)-2-(2'R,3'R-Dihydroxytricosanoylamino)- 8-octadecene-1,3,4-triol		$\begin{array}{ccc} R_1 & R_2 & R_3 \\ OH \\ \vdots \\ OH \\ OH \end{array}$	[25]
2	-	(2S,3S,4R,8E)-2-(2'R-Hydroxytetracosanoylamino)-10- octadecene-1,3,4-triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ & & $	[25]
3	-	(2S,3S,4R,8E)-2-(2'R-Hydroxytricosanoylamino)-10- octadecene-1,3,4-triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ \hline \\ OH & \\ \hline \\ OH & \\ \end{array}$	[25]
4	-	1-O-β-D-Glucopyranosyl-(2S,3S,4R,8E)-2-(2'R- Hydroxytetracosanoylamino)-8-octadecene-1,3,4- triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ & & & \\ OH & & \\ \end{array}$	[25]
5	-	1-O-β-D-Glucopyranosyl-(2S,3S,4R,8E)-2-(2'R- Hydroxytricosanoylamino)-8-octadecene-1,3,4-triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline$	[25]
6	-	1-O-β-D-Glucopyranosyl-(25,35,4R,8E)-2-(2'R- Hydroxyheptadecanoylamino)-8-octadecene-1,3,4- triol	Ceramide	$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ \hline \\ OH & & & \\ \end{array} \qquad \qquad$	[25]
7	-	1-O-β-D-Glucopyranosyl-(2S,3S,4R,8E)-2-(2'R- Hydroxydocosanoylamino)-8-octadecene-1,3,4-triol	R <sub>3</sub> O OH	$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & $	[25]
8	-	1-O-β-D-Glucopyranosyl-(25,35,4R,8E)-2-(2'R- Hydroxyhexadecanoylamino)-8-octadecene-1,3,4- triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ \hline \\ OH & & & \\ \end{array} \qquad \qquad$	[25]
9	-	1-O-β-D-Glucopyranosyl-(2S,3S,4R,8E)-2-(2'R- Hydroxyheptadecanoylamino)-8-octadecene-1,3,4- triol		$\begin{array}{cccc} R_1 & R_2 & R_3 \\ & & & \\ & & & \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline$	[25]
10	-	(2R)-Isookanin-4'-methoxy-7-Ο-β-D-glucopyranoside	Flavonoid	R <sub>1</sub> R <sub>2</sub> R <sub>3</sub> OGIC OCH <sub>3</sub> OH	[25]
11	-	(2S)-Isookanin-4'-methoxy-7-O-β-D-glucopyranoside	Ri Contraction Right	R1 R2 R3   OGIc OCH3 OH	[25]
12	-	(2R)-Isookanin-3'-methoxy-7-Ο-β-D-glucopyranoside		$\begin{array}{ccc} R_1 & R_2 & R_3 \\ \textbf{OGIc} & \textbf{OH} & \textbf{OCH}_3 \end{array}$	[25]

Annals of Pharmaceutical Sciences

13	-	(2S)-Isookanin-3'-methoxy-7-O-β-D-glucopyranoside		R1R2R3OGICOHOCH3	[25]
14	-	(2R)-7,8,3',4'-Tetrahydroxyflavone-3'-methoxy-7-Ο-β- D-glucopyranoside	-	R1 R2 R3   OH OH OH	[25]
15	-	(2S)-7,8,3',4'-Tetrahydroxyflavone-3'-methoxy-7-Ο-β- D-glucopyranoside	-	R <sub>1</sub> R <sub>2</sub> R <sub>3</sub> OH OH OH	[25]
16	-	E - 6 - O - β - D - G l u c o p y r a n o s y l - 6 , 7 , 3 ' , 4 ' - tetrahydroxyaurone	Flavonoid	R <sub>1</sub> R <sub>2</sub> OGIC OH	[25]
17	-	7-O-β-D-Glucopyranosyl-6,7,3',4'-tetrahydroxyaurone		R <sub>1</sub> R <sub>2</sub> OH OGIC	[25]
18	Maritimetin	3',5-Dihydroxy-4',6,7-trimethoxyflavone	°,	R <sub>1</sub> R <sub>2</sub> ОН ОН	[25]
19	-	E-4-O-(2''-O-diacetyl-6''-O-p-coumaroyl-β-D- glucopyranosyl)-p-coumaric acid	Phenylpropanoid		[25]
20	-	4-O-(6''-O-p-Sementoncoacyl-β-D-glucopyranosyl)-p- coumaric acid	Phenylpropanoid		[25]
21	Citrusin C	4'-Hydroxy-3,3',4',5,6,7-hexamethoxyflavone	Phenylpropanoid	R=H HOH H <sub>3</sub> CO HOH H <sub>OH</sub> OH R HOH H <sub>R</sub>	[25]
22	-	4-Allyl-2,6-dimethoxyphenyl glucoside	Phenylpropanoid	R=OCH <sub>3</sub> HoHOHHOHHOHR	[25]
23	Caffeic Acid	3-(3,4-Dihydroxyphenyl)-2-propenoic acid	Phenylpropanoid	HO HO U	[25]
24	Fumaric Acid	(2E)-But-2-enedioic acid	Aliphatic	HO	[25]
25	Uracil	2,4-Dioxopyrimidine	Pyrimidine		[25]
26	β-Sitosterol	(3β)-Stigmast-5-en-3-ol	Steroid		[25]
27	Daucosterol	(3β)-Stigmast-5-en-3-yl β-D-glucopyranoside	Steroid		[25]

28	Stigmasterol	(22E)-Stigmasta-5,22-dien-3β-ol	Steroid	HO HO	[25]
29	Stigmasterol- β-D- glucopyranoside	(3β,22E)-Stigmasta-5,22-dien-3-yl β-D- glucopyranoside	Steroid		[25]
30	Friedelin	Friedo-olean-3-one	Triterpenoid		[25]
31	Stearic Acid	Octadecanoic acid	Aliphatic	Он	[25]
32	Hexadecanol	Hexadecan-1-ol	Aliphatic	······	[25]
33	n-Heneicosane	Heneicosane	Aliphatic	~~~~~~	[25]
34	-	(5E)-Trideca-1,5-dien-7,9,11-triyne-3,4-diol-4-O-β-D- glucopyranoside	Polyacetylene		[25]
35	Linoleic Acid	cis, cis-9,12-Octadecadienoic acid	Fatty acid (Polyunsaturated omega-6 fatty acid)	HO 1 9 12	[20]
36	Dehydroabietic Acid	4aS,10aS)-8,8-Dimethyl-4,4a,9,10,10a,10b-hexahy- dro-7-oxophenanthrene-2-carboxylic acid	Diterpenoid		[20]
37	Bidenoside C	(2Z)-deca-2-ene-4,6-diyn-1-O-β-D-glucopyranoside	Acetylenic Glucoside	QIc-0-CH2 <sup>-</sup> -ĊH2-ĊH2-Ċ≡Ċ-Ċ≡Ċ-Ċ	[26]
38	Bidenoside D	(2E)-deca-2-ene-4,6-diyn-3,10-dihydroxy-1-O-β-D- glucopyranoside	Acetylenic Glucoside		[26]

39	Bidenoside F	5,7-dihydroxy-2-(4-hydroxyphenyl)-6-methoxy-8-(2,4- dihydroxyphenyl)-4H-1-benzopyran-4-one 7-O-(3',6'- di-O-acetyl)-β-D-glucopyranoside	Flavanone glucoside	$\begin{array}{c} 6^{3} & OAc \\ 5^{5} & O \\ 4^{7} OAc \\ 4^{7} OAc \\ 0Ac \\ 0H \\ 0H \\ 0H \\ H \\ 0H \\ H \\ 0 \\ 0$	[27]
40	Bidenoside G	3,4,2',4'-tetrahydroxychalcone-4'-O-(6''-O-acetyl)-β- D-glucopyranoside	Chalcone glucoside	$\begin{array}{c} & & & & & & & & & & & & & & & & & & &$	[27]
41	(2S) (5E,11E)- Tridecadiene- 7,9-diyne-1,2,13- triol	(2S,5E,11E)-Trideca-5,11-dien-7,9-diyn-1,2,13-triol	Polyacetylene	HO 13 12 10 9 8 7 6 5 4 3 0H 2 10 0H	[28]
42	(6E,12E)-3-oxo- tetradecadiene- 8,10-diyne- 14-hydroxyl- 1-O-β-D- glucopyranoside	(6E,12E)-3-Oxo-tetradeca-6,12-dien-8,10-diyn-14-yl β-D-glucopyranoside	Polyacetylene	HO 14 12 12 16 10 9 8 7 6 3 2 10 10 9 8 7 6 3 2 10	[28]
43	(6E,12E)-Tetra- decadiene-8,10- diyne-1,3,14- triol	(6E,12E)-Tetradeca-6,12-dien-8,10-diyn-1,3,14-triol	Polyacetylene	НО	[28]
44	(6E,12E)- Tetradecadiene- 8,10-diyne-1,14- diol-3-Ο-β-D- glucopyranoside	(6E,12E)-Tetradeca-6,12-dien-8,10-diyn-1,14-diol-3- Ο-β-D-glucopyranoside	Polyacetylene	$\frac{HO}{14} \underbrace{\frac{13}{12}}_{R=Glucose} \underbrace{\frac{11}{10} \cdot 9 \cdot 8}_{7 \times 6} \underbrace{\frac{9}{5} \cdot 4 \cdot 5}_{7 \times 6} \underbrace{OR}_{3 \times 2}_{OH}$	[28]
45	(3E,11E)-Tride- cadiene-6,8,10- atriyne-1,2,13- triol	(3E,11E)-Trideca-3,11-dien-6,8,10-triyn-1,2,13-triol	Polyacetylene	$R=H^{HO} \xrightarrow{12} 10 \xrightarrow{9} 8 \xrightarrow{7} 6 \xrightarrow{5} 4 \xrightarrow{3} 2^{OR}$	[28]
46	(3E,11E)- Tridecadiene- 6,8,10-atriyne- 1,13-diol- 2-O-β-D- glucopyranoside	(3E,11E)-Trideca-3,11-dien-6,8,10-triyn-1,13-diol-2- Ο-β-D-glucopyranoside	Polyacetylene	$\frac{HO}{13} \underbrace{10_{9}}_{H1} \underbrace{9 \times \frac{7}{6} \times \frac{6}{5}}_{4 \times \frac{3}{2}} \underbrace{9 \times \frac{8}{7}}_{2 \times \frac{6}{5}} \underbrace{4 \times \frac{3}{2}}_{0 \text{H}} OR}_{R=Glucose}$	[28]

47	3,6,8-Trichloro- 5,7,3',4'-tetrahy- droxyflavone	3,6,8-Trichloro-5,7,3',4'-tetrahydroxy-2-phenyl-4H-1- benzopyran-4-one	Chlorinated Flavonoid	$\begin{array}{c} OH \\ OH \\ HO \\ 7 \\ HO \\ 7 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10 $	[29]
48	p-Coumaroyl glucoside	1-O-(E)-4-hydroxycinnamoyl-6-O-(E)-4-hydroxycinna- moyl-2,3-di-O-acetyl-β-D-glucopyranose	Phenylpropanoid Glycoside	HO =	[29]
49	Apigenin- 7-O-β-D- glucopyranoside	5,7-Dihydroxy-2-(4-hydroxyphenyl)-4H-chromen-4- one 7-O-β-D-glucopyranoside	Flavone Glycoside	$\begin{array}{c} HO\\HO\\HO\\HO\\3^{*} & 2^{*}OH\\HO\\HO\\3^{*} & 2^{*}OH\\H\\0\end{array} \xrightarrow{(1)}{} 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 $	[29]
50	Luteolin	5,7,3',4'-Tetrahydroxyflavone	Flavonoid	HO HC HC CH	[29,30]
51	5,7,3',4'-Tetra- hydroxy-6-me- thoxyflavone	6-Methoxy-5,7,3',4'-tetrahydroxy-2-phenyl-4H-1-ben- zopyran-4-one	Flavonoid	HO HO OH COOH	[29]
52	3',4'-Dihydroxy- flavone	3',4'-Dihydroxy-2-phenyl-4H-1-benzopyran-4-one	Flavonoid	HO OH OH	[29]
53	Quercetin	2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-4H-1-ben- zopyran-4-one	Flavonol	но он он он он он он	[29]
54	Kaempferol	3,5,7-Trihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopy- ran-4-one	Flavonol	HO OH OH OH	[29]
55	Isorhamnetin	3,5,7-Trihydroxy-2-(4-hydroxy-3-methoxyphenyl)-4H- 1-benzopyran-4-one	Flavonol	HO OH	[29]
56	Rutin	5,7,3',4'-Tetrahydroxyflavonol-3-rutinoside	Flavonol Glycoside	HO OH O	[29,30]

57	Quercitrin	5,7,3',4'-Tetrahydroxyflavonol-3-rhamnoside	Flavonol Glycoside	HO O OH OH O HO OC HO OC HO OC HO OC	[29,30]
58	Isoquercitrin	5,7,3',4'-Tetrahydroxyflavonol-3-glucoside	Flavonol Glycoside	HO OH OH OH OH OH OH OHOHOH	[25,29]
59	Luteolin- 7-O-β-D- glucopyranoside	5,7,3',4'-Tetrahydroxyflavone 7-O-β-D- glucopyranoside	Flavone Glycoside		[29]
60	Apigenin	5,7-Dihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran- 4-one	Flavone	HO O OH OH O	[29]
61	Acacetin	5,7-Dihydroxy-4'-methoxyflavone	Flavone	HO OCH3 OH O	[29]
62	Diosmetin	5,7,3'-Trihydroxy-4'-methoxyflavone	Flavone	HO OH OH OH OH	[29]
63	Chrysoeriol	5,7,3'-Trihydroxy-4'-methoxyflavone	Flavone	HO O O O O O H3 O H	[29]
64	Hispidulin	5,7,4'-Trihydroxy-6-methoxyflavone	Flavone	HO O O HO O HO	[29]

65	Tricin	5,7,4'-Trihydroxy-3',5'-dimethoxyflavone	Flavone	HO OCH <sub>3</sub> OH OCH <sub>3</sub> OH	[29]
66	Caffeic acid	(E)-3-(3,4-Dihydroxyphenyl)prop-2-enoic acid	Phenolic Acid	НО ОН	[29]
67	Chlorogenic acid	(1S,3R,4R,5R)-3-{[(2Z)-2-(3,4-Dihydroxyphenyl)ethe- noyl]oxy}-1,4,5-trihydroxycyclohexane-1-carboxylic acid	Phenolic Acid	HO, CO <sub>2</sub> H HO <sup>11</sup> O HO <sup>11</sup> O HO O HO HO HO HO HO HO HO HO HO HO HO	[29]
68	Ferulic acid	(E)-3-(4-Hydroxy-3-methoxyphenyl)prop-2-enoic acid	Phenolic Acid	CH <sub>3</sub> O HO	[29]
69	p-Coumaric acid	(E)-3-(4-Hydroxyphenyl)prop-2-enoic acid	Phenolic Acid	но	[29]
70	Gallic acid	3,4,5-Trihydroxybenzoic acid	Phenolic Acid	но об он	[29,30]
71	Vanillic acid	4-Hydroxy-3-methoxybenzoic acid	Phenolic Acid	O OH OCH <sub>3</sub>	[29]
72	Syringic acid	4-Hydroxy-3,5-dimethoxybenzoic acid	Phenolic Acid	О ОН	[29]
73	Protocatechuic acid	3,4-Dihydroxybenzoic acid	Phenolic Acid	О ОН ОН ОН	[29]

74	3,6,8-Trichloro- 5,7,3',4'-tetrahy- droxyflavone	3,6,8-Trichloro-5,7,3',4'-tetrahydroxy-2-phenyl-4H-1- benzopyran-4-one	Chlorinated Flavonoid		[29]
75	Hyperoside	3-(β-D-Galactopyranosyloxy)-3',4',5,7- tetrahydroxyflavone	flavonol glycoside		[25,30]
76	Protocatechuic acid	3,4-Dihydroxybenzoic acid	Phenolic	ОНОНОН	[30]
77	Bipinnat poly- acetylenicloside	-	-		[30]
78		9-oxo-(10E, 12E) octadecadienoic acid	Oxo fatty acid	CH OF CHARGE CHA	[31]
79		8-oxo-(9E, 11E)-octadecadienoic acid	Oxo fatty acid		[31]
80	Sterigmatocystin	(3aR,12cS)-8-Hydroxy-6-methoxy-3a,12c-dihydro-7H- furo[3',2':4,5]furo[2,3-c]xanthen-7-one	Coumarin derivative (Furanocoumarin)		[31]
81	Aflatoxin B1	(6aR,9aS)-4-Methoxy-2,3,6a,9a- tetrahydrocyclopenta[c]furo[3',2':4,5]furo[2,3-h][1] benzopyran-1,11-dione	Coumarin derivative (Furanocoumarin)	H H H H O CH <sub>3</sub>	[31]

## **Pharmacological Activity**

## Antibacterial activity

Antibiotic resistance has emerged as a major worldwide health concern in recent years, particularly in agricultural settings. In the current investigation, Campylobacter jejuni revealed 100% resistance to Ampicillin, which is consistent with earlier studies that have documented amoxicillin-resistant commensal Campylobacter. Amoxicillin alone was unsuccessful, while Co-amoxiclav revealed great activity against amoxicillinresistant bacteria [32]. This underscores the importance of judicious antibiotic use in the cattle industry, as the emergence of high resistance necessitates careful management [33]. Similarly, Klebsiella species demonstrated resistance to ampicillin while remaining vulnerable to cephalosporins, aminoglycosides, and quinolones [34].

B. bipinnata demonstrated significant efficacy against various tested bacteria, particularly those isolated from cattle waste. This suggests its potential in preventing cattle infections caused by these organisms [35]. B. bipinnata exhibits notable antibacterial activity due to compounds such as caffeic acid [36], chlorogenic acid [37], ferulic acid [38], gallic acid [39], and quercetin [40]. B. bipinnata contains two significant antibacterial substances: 16-pregnenolone as well as 9-octadecenoic acid (Z)-methyl ester.16-Pregnenolone, a steroidal molecule, showed excellent antibacterial activity against Staphylococcus aureus, with a MIC50 of 72  $\mu$ g/mL. In contrast, 9-Octadecenoic acid (Z)-methyl ester had a larger MIC50 value of nearly 250 µg/ mL, suggesting decreased potency. Furthermore, the pharmacological efficiency of 16-pregnenolone is supported by its amphiphilic nature, which allows it to break bacterial membranes by targeting particular membrane components and thereby exerting its bactericidal effects [41].

#### **Antitumor Activity**

Human hepatocellular carcinoma (HepG2) and cervical cancer (Hela) cell lines were significantly inhibited by extracts of B. bipinnata in vitro. The MTT test was used to measure the extract's effect on cell growth at different concentrations, which led to the calculation of IC50 values. According to our findings, HepG2 and Hela cells are significantly inhibited, with the greatest suppression shown after 48 hours of therapy. One has an IC50 value of 14.80  $\mu$ g/mL while the other is 13.50  $\mu$ g/mL. The extract of B. bipinnata showed promising inhibitory effects on many cell lines, suggesting that it might be used in future studies [42]. In previous research, phytochemicals found in B. bipinnata were shown to have anticancer effects. Plant compounds that may have medicinal significance include maritimetin, citrusin C, fumaric acid, apigenin, luteolin, quercetin, kaempferol, dehydroabietic acid, and caffeic acid. The plant's effectiveness against cancer, and specifically U14 cells from cervical carcinoma, has been brought to light in recent investigations. The MTT test was used in in vitro research to show that B. bipinnata extract inhibits U14 cell growth in a way that is dependent on both dose and time. At 80µg/L, the inhibition rate was 70.44%. Extensive tumor growth inhibition and life extension advantages were shown in in vivo testing conducted on mice with U14 solid and ascites tumors. With a tumor suppression rate of 49.13% in high-dose groups and a life extension rate of 63.63% for ascites tumors, the extract clearly shows promise as an adjuvant therapy for cancer diagnosis and treatment [43]. One of the most well-known flavonoids found in B. bipinnata, isoquercitrin, has strong pharmacological effects, especially in the fight against liver cancer. Isoquercitrin has shown a dose- and time-dependent ability to inhibit the proliferation of HepG2 and Hep3B, two human liver cancer cell lines. At concentrations between 100 and 800  $\mu$ M, there was a significant decrease in cell viability. A concentration-dependent increase in apoptotic cells was shown using Annexin V-FITC/PI double labelling flow cytometry, demonstrating that the chemical causes apoptosis. In addition to its antiproliferative effects, isoquercitrin causes cell cycle arrest in the G1 phase. The activation of caspases -3, -8, and -9 isoquercitrin's apoptotic action, suggesting that both intrinsic and extrinsic mechanisms are involved. And by increasing JNK phosphorylation and lowering ERK and p38MAPK phosphorylation, isoquercitrin modifies the MAPK transmission pathway. There is a reduction in the expression of protein kinase C (PKC), which is essential for the continued growth and survival of cells. Isoquercitrin significantly inhibited the growth of transplanted liver tumors in nude mice, lending credence to these results in in vivo research. The many paths that isoquercitrin can follow demonstrate its therapeutic potential and point to *B. bipinnata* as a plant that could provide bioactive compounds with strong anticancer effects [44].

# Antidiarrhoeal Activity

B. bipinnata methanol extract showed significant antidiarrheal effectiveness in many pharmacological studies. B. bipinnata significantly reduced castor oil-induced diarrhoea in rats, according to a study that compared the antidiarrheal effects of several plant extracts. The number of soft faecal pellets was considerably reduced in the group that received the extract at dosages of 200 mg/kg and 400 mg/kg, in comparison to the control group. In addition, while measuring gastrointestinal motility with a charcoal meal, the longer the charcoal rods stayed in the test tube after consuming a higher dosage of B. bipinnata extract, suggesting less transit time through the intestines. To further demonstrate its antidiarrheal properties, the methanol extract dose-dependently reduced motility in isolated rabbit duodenum. The presence of tannins, flavonoids, carbohydrates, lactones, unsaturated sterols/triterpenes, and proteins/amino acids was shown by phytochemical analysis. These compounds are likely responsible for the antidiarrheal effects observed [45].

# Anti-inflammatory Activity

Among the several pharmacological effects of B. bipinnata are its cytotoxic and anti-inflammatory properties. Flavonoids, sesquiterpene lactones, polyacetylenes, and other bioactive compounds are present in the plant and help explain its medicinal effects. Along with four other polyacetylenes that were previously reported, two more, (2S) (5E,11E)-tridecadiene-7,9diyne-1,2,13-triol and (6E,12E)-3-oxo-tetradecadiene-8,10diyne-14-hydroxyl-1-O-β-D-glucopyranoside, have demonstrated an impressive anti-inflammatory impact, joining four previously identified polyacetylenes. These substances block macrophage cells from responding to lipopolysaccharide (LPS) by releasing TNF- $\alpha$  and IL-1 $\beta$ , which means they have anti-inflammatory qualities. Even though these polyacetylenes were quite effective in reducing inflammation, they showed very little cytotoxicity when tested against human cancers of the cervical region (HeLa), human carcinomas of the liver (HepG-2), and human cancerous breast cells (MCF-7). Extensive research has shown that HUVECs, when exposed to serum from individuals with Henoch-Schönlein purpura (HSP), release many cytokines that promote inflammation, including Nitric Oxide (NO), interleukin-8 (IL-8), and Tumour Necrosis Factor-Alpha (TNF- $\alpha$ ) [46]. Its significant anti-inflammatory activity is mainly due to the total flavonoid content (TFB) of B. bipinnata. Research has demonstrated that when stimulated with sera from Henoch-Schönlein purpura (HSP) patients, human umbilical vein endothelial cells (HUVECs) produce a number of pro-inflammatory cytokines, such as Nitric Oxide (NO), interleukin-8 (IL-8), and tumour necrosis factor-alpha (TNF- $\alpha$ ). However, when treated with TFB, these cytokines are effectively inhibited. In addition, two important mediators of the inflammatory response, nuclear factor-kappa B (NF-kB) and fractalkine, are inhibited in their mRNA and protein production by TFB. According to these studies, TFB decreases inflammation by blocking NF-kB signalling pathways, which leads to cytokine production [47]. B. bipinnata flavonoids (BBTF) may have anti-inflammatory effects due to their capacity to reduce endothelial cell production of inflammatory mediators [48].

## **Antioxidant Activity**

Recent pharmacological studies have highlighted the plant's diverse bioactivities, attributing its therapeutic potential primarily to its rich flavonoid content. The total flavonoids from *B. bipinnata* (BBTF) have demonstrated significant antioxidant activities [48].

B. bipinnata, possess antioxidative effects. The antioxidative properties of B. bipinnata are attributed to its ability to inhibit linoleic acid peroxidation and scavenge DPPH radicals. Five major flavonoids have been isolated from the plant's floral ethanol extract: sulfuretin, butein, 7,8,3',4' tetrahydroxyflavanone, maritimetin, and okanin. Sulfuretin and butein are highly effective chemicals that show significant antioxidative activity. These results validate the traditional usage of B. bipinnata in the management of oxidative stress by highlighting its potential as a natural source of antioxidants [49]. It has been demonstrated that the flavonoid-rich extract of *B. bipinnata* shields β-cells of the pancreas against oxidative stress-triggered apoptosis. The effect of the extract (BBTF) on INS-1 cells exposed to hydrogen peroxide (H2O2), a common oxidative stressor, was examined in this work. BBTF pretreatment increased β-cell viability, lowered ROS production, and decreased apoptosis. The underlying processes were unveiled by the modulation of apoptosis-related proteins: BBTF reduced levels of the pro-apoptotic protein Bax and increased levels of the anti-apoptotic protein Bcl-2. It also reduced the phosphorylation of stress-related kinases (JNK, ERK, and p38) and lowered the production of death receptorrelated proteins (Fas and FasL), resulting in a reduction in caspase-8, caspase-9, and caspase-3 activity. The ability of the B. bipinnata extract, which is rich in flavonoids, to protect pancreatic  $\beta$ -cells from oxidative damage and ensure their survival and function might make it useful in managing diabetes [50].

# Antidiabetic Activity

The phenolic components of *B. bipinnata* are primarily responsible for the significant anti-diabetic effects of its ethyl acetate extract. The enzyme  $\alpha$ -amylase, which converts starch to glucose, is inhibited by isookanin. Isookanin can regulate postprandial blood glucose levels by inhibiting  $\alpha$ -amylase, which makes it a useful treatment for diabetes. This study highlights the medicinal potential of *B. bipinnata*, opening the door for its application in the creation of functional foods and pharmaceutical formulations intended to manage diabetes and its related symptoms [51]. *B. bipinnata* (BBTF) flavonoids in their whole form have a regulating impact on insulin resistance through the PI3K/AKT1/GLUT4 signaling pathway [48].

# Antihyperlipidemic Effect

*B. bipinnata* extracts have been proven to significantly lowdensity lipoprotein cholesterol (LDL-C) and lower blood total cholesterol (also known as TC), while increasing HDL-C. Furthermore, these extracts increase antioxidant enzyme activity, such as superoxide dismutase (SOD), while decreasing oxidative stress indicators including malonaldehyde (MDA) and Nitric Oxide (NO). The underlying processes involve modification of the PPAR signalling pathway, which is critical for regulating lipid metabolism. These findings imply that *B. bipinnata* may be an effective natural therapy for treating hyperlipidemia, with potential uses in decreasing the risk of cardiovascular disease [52]. *B.* 

bipinnata exhibits significant antihyperlipidemic activity, making it a promising candidate for managing hyperlipidemia, a major risk factor for cardiovascular diseases. Work has indicated that the whole extract of B. bipinnata is capable of efficiently controlling blood lipid levels. The extract significantly reduces blood levels of Triglycerides (TG), lower-density lipoprotein cholesterol (LDL-C), and Total Cholesterol (TC) in hyperlipidemic rat models, while increasing HDL-C levels. This lipid-regulating effect is primarily attributed to several active compounds, including gallic acid, protocatechuic acid, rutin, hyperoside, bipinnata polyacetylenicloside, luteolin, and guercetin. These compounds exhibit diverse chemical structures and are predominantly distributed in the liver, which is a key organ in lipid metabolism . Additionally, molecular docking studies have identified significant binding affinities between these compounds and target proteins such as HMGCR, NR3C1, CYP1A2, RXRA, CES1, HSD11B1, and CYP1A1, suggesting their potential mechanisms in modulating lipid metabolism and exerting antihyperlipidemic effects [30].

# Antiarthritic activity

Total flavonoids from *B. bipinnata* (TFB) have been proven in trials to alleviate adjuvant-induced arthritis in rats by reducing synovial hyperplasia, inflammatory cell infiltration, and cartilage breakdown. TFB inhibits the synthesis of cytokines that trigger inflammation such as IL-1 $\beta$ , TNF- $\alpha$ , and IL-6. TFB also causes synovial apoptosis, as seen by an increase in TUNEL-positive cells, DNA fragmentation, and caspase-3 activity. These findings demonstrate that TFB from *B. bipinnata* may offer a therapeutic method for treating inflammatory illnesses like RA due to its dual anti-inflammatory and pro-apoptotic activity [53].

## Hepatoprotective Effect

*B. bipinnata's* total flavonoids (TFB) showed substantial hepatoprotective benefits. According to experimental research, TFB has strong antioxidant capabilities that play an important role in preventing oxidative stress-induced liver damage. TFB treatment has been demonstrated to diminish high liver and spleen indices, serum transaminase levels, and liver fibrosis indicators including hyaluronic acid and type III procollagen. Lipid peroxidation is decreased because TFB increases levels of glutathione peroxidase and superoxide dismutase, two antioxidant enzymes. TFB protects against liver fibrosis by inhibiting the expression of NF-κB, α-SMA, and TGF-β1 genes, all of which play important roles in the fibrogenic process. These findings indicate that TFB might be a viable therapeutic drug in the treatment of liver fibrosis by reducing oxidative stress and regulating important fibrogenic pathways [54].

# Neuropharmacological Activity

The pharmacological activity of *B. bipinnata*, often called Spanish needles or beggar's ticks, is substantial and includes neuroprotective properties. Research on rats has shown that total flavones from Bidens bipinnata L. (TFB) can prevent brain harm following experimental intracerebral hemorrhage. Reducing cerebral edema, improving microcirculation, and increasing antioxidant defenses are the primary determinants of this neuroprotective impact. Malondialdehyde (MDA) and Nitric Oxide (NO), two markers of oxidative stress and inflammation, are decreased by TFB treatment, while Superoxide Dismutase (SOD) activity is greatly increased. By reducing oxidative damage and enhancing neurological outcomes, these flavones forestall lipid peroxidation and NO production [55].

## Conclusion

B. bipinnata is a tall annual plant that grows in low-temperature tropical locations all over the world. It has green leaves, yellow or white blooms, and brownish-blackish seeds. You may find it growing near roadside. B. bipinnata is a very nutritious food source that has an extensive record of usage in conventional healthcare. Research on B. bipinnata has been conducted in several disciplines, including botany, ethnomedicine, pharmacology, and phytochemistry. The wide variety of bioactive substances found in B. bipinnata, including tannins, terpenoids, vaginitis, candidiasis, stomachaches, dysentery, vaginitis, and bronchial illnesses, as well as many others, have long been utilized for medicinal purposes. Thanks to these phytoconstituents, it can effectively cure a variety of diseases and conditions, including as cancer, inflammation, diabetes, bacterial infections, gastrointestinal issues, and cardiovascular maladies. The therapeutic uses of B. bipinnata are highlighted, and its medicinal potential is examined in connection to its recognized phytochemicals. However, due to the possibility of hypoglycemia, hypotension, and allergic responses, it should be used with caution. Complete validation and exploitation of B. bipinnata's therapeutic value requires more study and clinical studies.

#### **Future Perspectives**

To enhance its use in contemporary medicine, future studies on *B. bipinnata* should concentrate on a few important areas. First things first: we need more comprehensive phytochemical research to identify and define the active ingredients that are therapeutically useful. Secondly, in order to have a better understanding of the medications' metabolism, excretion, distribution, and absorption, thorough pharmacokinetic and pharmacodynamic investigations should be carried out. To conclude, the effectiveness and safety of B. bipinnata in human populations can only be ascertained by conducting large-scale clinical studies. More research into its synergistic effects with other medicinal herbs or traditional pharmaceuticals could lead to exciting new possibilities in combination treatment. Last but not least, new treatments utilizing B. bipinnata can be developed by combining traditional knowledge with state-of-the-art scientific research.

#### References

- Soni P, Siddiqui A A, Dwivedi J, Soni V. Pharmacological properties of Datura stramonium L. as a potential medicinal tree: an overview. Asian Pacific journal of tropical biomedicine. 2012; 2(12): 1002-1008. https://doi.org/10.1016/S2221-1691(13)60014-3.
- Asigbaase M, Adusu D, Anaba L, Abugre S, Kang-Milung S, et al. Conservation and economic benefits of medicinal plants: Insights from forest-fringe communities of Southwestern Ghana. Trees, Forests and People. 2023; 14. https://doi.org/10.1016/j. tfp.2023.100462.
- Elsanhoty RM, Soliman MSM, Khidr YA, Hassan GOO, Hassan ARA, et al. Pharmacological Activities and Characterization of Phenolic and Flavonoid Compounds in Solenostemma argel Extract. Molecules. 2022; 27: 8118. https://doi.org/10.3390/molecules27238118.
- Mehmood A, Javid S, Khan M F, Ahmad K S, Mustafa A. In vitro total phenolics, total flavonoids, antioxidant and antibacterial activities of selected medicinal plants using different solvent systems. BMC chemistry. 2022; 16(1): 64. https://doi.org/10.1186/ s13065-022-00858-2.



Figure 2: Phytochemical & pharmacological profile of *B.bipinnata* 

- SIRBU, CULITA, OPREA, ADRIAN. New Alien Species for the Flora of Romania: Bidens bipinnata L. (Asteraceae), Turkish Journal of Botany. 2008; 32(3): 7. https://journals.tubitak.gov.tr/botany/ vol32/iss3/7.
- Zahara K, Bibi Y, Tabassum S, Mudrikah Bashir T, Haider S, et al. A review on pharmacological properties of Bidens biternata: A potential nutraceutical. In Asian Pacific Journal of Tropical Disease. 2015; 5(8): 595-599. Elsevier B.V. https://doi.org/10.1016/ S2222-1808(15)60894-5.
- Bernal R, G Galeano, A Rodríguez, H Sarmiento y, M Gutiérrez. Nombres Comunes de las Plantas de Colombia. 2017. http:// www.biovirtual.unal.edu.co/nombrescomunes.
- Bartolome A P, Villaseñor I M, Yang W C. Bidens pilosa L. (Asteraceae): Botanical Properties, Traditional Uses, Phytochemistry, and Pharmacology. Evidence-based complementary and alternative medicine: eCAM. 2013; 340215. https://doi.org/10.1155/2013/340215.
- Florida State University Robert K. Godfrey Herbarium database. Tallahassee (FL): Florida State University; Collectors: Bian Tan, Loran C. Anderson, Ed Keppner, Lisa Keppner, Richard S. Mitchell, R.K. Godfrey, R. Kral, Wilson Baker, R. Komarek, Andre F. Clewell, R.A. Norris, and Andre F. Clewell. States and Counties: Florida: Columbia, Wakulla, Bay, Calhoun, Leon, Franklin, Jackson, Jefferson, and Liberty. Georgia: Grady. 2014. http://herbarium.bio. fsu.edu.
- 10. Bidens bipinnata. In Wikipedia. 2024. https://en.wikipedia.org/ wiki/Bidens bipinnata
- Gurucharan singh, flowers of india, Spanish Needles. 2024. https://www.flowersofindia.net/catalog/slides/Spanish%20 Needles.html.

- 12. Vladimirov V, Petrova AS. Two alien species of Bidens (Asteraceae) new to the Bulgarian flora. Phytologia Balcanica. 2009; 15. https://www.researchgate.net/publication/239532186.
- 13. USDA. Agricultural Research Service, National Plant Germplasm System. Germplasm Resources Information Network (GRIN Taxonomy). National Germplasm Resources Laboratory, Beltsville, Maryland. 2024. https://npgsweb.ars-grin.gov/gringlobal/taxon/taxonomydetail?id=104072.
- 14. Parr C S, N Wilson, P Leary, K S Schulz, K Lans, et al. The Encyclopedia of Life v2: Providing Global Access to Knowledge about Life on Earth. Biodiversity Data Journal. 2014; 2: e1079. http:// doi.org/10.3897/BDJ.2.e1079.
- Andrade-Neto V F, Brandão M G, Oliveira F Q, Casali V W, Njaine B, et al. Antimalarial activity of Bidens pilosa L. (Asteraceae) ethanol extracts from wild plants collected in various localities or plants cultivated in humus soil. Phytotherapy research: PTR. 2004; 18(8): 634-639. https://doi.org/10.1002/ptr.1510.
- Solanki Y, Kumar N, Meena A, Singh j, Kotiya A, et al. (Asteraceae): A new species record to Rajasthan state, India; IJB. 2023; 23(5): 54-59 https://innspub.net/bidens-bipinnata-l-asteraceae-a-new-species-record-to-rajasthan-state-india/.
- Kil J S, Son Y, Cheong YK, Kim N H, Jeong H J, et al. Okanin, a chalcone found in the genus Bidens, and 3-penten-2-one inhibit inducible nitric oxide synthase expression via heme oxygenase-1 induction in RAW264.7 macrophages activated with lipopolysaccharide. Journal of clinical biochemistry and nutrition. 2012; 50(1): 53-58. https://doi.org/10.3164/jcbn.11-30.
- AlSheikh H M A, Sultan I, Kumar V, Rather I A, Al-Sheikh H, et al. Plant-Based Phytochemicals as Possible Alternative to Antibiotics in Combating Bacterial Drug Resistance. Antibiotics (Basel, Switzerland). 2020; 9(8): 480. https://doi.org/10.3390/antibiotics9080480.
- Bseiso E A, Nasr M, Sammour O, Abd El Gawad N A. Recent advances in topical formulation carriers of antifungal agents. Indian journal of dermatology, venereology and leprology. 2015; 81(5): 457-463. https://doi.org/10.4103/0378-6323.162328.
- Zahara K, Bibi Y, Masood S, Nisa S, Qayyum A, et al. Using HPLC-DAD and GC-MS Analysis Isolation and Identification of Anticandida Compounds from Gui Zhen Cao Herbs (Genus Bidens): An Important Chinese Medicinal Formulation. Molecules (Basel, Switzerland). 2021; 26(19): 5820. https://doi.org/10.3390/molecules26195820.
- 21. Mohammadi S, Keshteli A H, Saneei P, Afshar H, Esmaillzadeh A, et al. The Relationship Between Linoleic Acid Intake and Psychological Disorders in Adults. Frontiers in nutrition. 2022; 9: 841282. https://doi.org/10.3389/fnut.2022.841282.
- 22. Lin C J, Lai C K, Kao M C, Wu L T, Lo U G, et al. Impact of cholesterol on disease progression. BioMedicine. 2015; 5(2): 7. https:// doi.org/10.7603/s40681-015-0007-8.
- 23. Jung S W, Thamphiwatana S, Zhang L, Obonyo M. Mechanism of antibacterial activity of liposomal linolenic acid against Helicobacter pylori. PloS one. 2015; 10(3): e0116519. https://doi.org/10.1371/journal.pone.0116519.
- 24. Hao M, Xu J, Wen H, Du J, Zhang S, et al. Recent Advances on Biological Activities and Structural Modifications of Dehydroabietic Acid. Toxins. 2022; 14(9): 632. https://doi.org/10.3390/toxins14090632.
- 25. Hu HM, Bai SM, Chen LJ, Hu WY, Chen G. Chemical constituents from Bidens bipinnata Linn. Biochem Syst Ecol. 2018; 79: 44-9. https://doi.org/10.1016/j.bse.2018.05.005.
- 26. Li S, Kuang H X, Okada Y, Okuyama T. New acetylenic glucosides

from Bidens bipinnata LINNE. Chemical & pharmaceutical bulletin. 2004; 52(4): 439-440. https://doi.org/10.1248/cpb.52.439.

- Li S, Kuang H X, Okada Y, Okuyama T. New flavanone and chalcone glucosides from Bidens bipinnata Linn. Journal of Asian natural products research. 2005; 7(1): 67-70. https://doi.org/1 0.1080/10286020310001617147 28.
- 28. Wang X, et al. Polyacetylenes from Bidens bipinnata L. and their biological activities. Phytochem. Lett. 2013. http://dx.doi. org/10.1016/j.phytol.2013.12.002.
- 29. Yang X W, Huang M Z, Jin YS, Sun L N, Song Y, et al. Phenolics from Bidens bipinnata and their amylase inhibitory properties. Fitoterapia. 2012; 83(7): 1169-1175. https://doi.org/10.1016/j. fitote.2012.07.005.
- Zhuang G, Wang Y Q, Li S J, Jiang X, Wang X Y. Tissue distribution and molecular docking research on the active components of Bidens bipinnata L. against hyperlipidemia. Biomedical chromatography: BMC. 2021; 35(4): e5026. https://doi.org/10.1002/ bmc.5026
- 31. Abdou R, Shaker K. Bioactive Metabolites of the Endophyte Khuskia Oryzae Isolated from the Medicinal Plant Bidens Bipinnata. In Asian Journal of Pharmacy and Life Science. 2013; 3(3). www.ajpls.com.
- Elviss N C, Williams L K, Jørgensen F, Chisholm S A, Lawson A J, et al. Amoxicillin therapy of poultry flocks: effect upon the selection of amoxicillin-resistant commensal Campylobacter spp. The Journal of antimicrobial chemotherapy. 2009; 64(4): 702-711. https://doi.org/10.1093/jac/dkp277.
- Gharbi M, Béjaoui A, Ben Hamda C, Jouini A, Ghedira K, et al. Prevalence and Antibiotic Resistance Patterns of Campylobacter spp. Isolated from Broiler Chickens in the North of Tunisia. BioMed research international. 2018; 7943786. https://doi. org/10.1155/2018/7943786.
- Stock I, Wiedemann B. Natural antibiotic susceptibility of Klebsiella pneumoniae, K. oxytoca, K. planticola, K. ornithinolytica and K. terrigena strains. Journal of medical microbiology. 2001; 50(5): 396-406. https://doi.org/10.1099/0022-1317-50-5-396.
- 35. Ahmad B, Khan A A, Maria H, Zaman S, Gul A, et al. Antibacterial Effect Of Bidens Bipinnata L. Extract against Selected Bacterial Strains. Repéré à. 2023. http://xisdxjxsu.asia.
- Park M Y, Kang D H. Antibacterial Activity of Caffeic Acid Combined with UV-A Light against Escherichia coli O157: H7, Salmonella enterica Serovar Typhimurium, and Listeria monocytogenes. Applied and environmental microbiology. 2021; 87(15): e0063121. https://doi.org/10.1128/AEM.00631-21.
- Chen K, Peng C, Chi F, Yu C, Yang Q, et al. Antibacterial and Antibiofilm Activities of Chlorogenic Acid against Yersinia enterocolitica. Frontiers in microbiology. 2022; 13: 885092. https://doi. org/10.3389/fmicb.2022.885092.
- Song W, Xin J, Yu C, Xia C, Pan Y. Alkyl ferulic acid esters: Evaluating their structure and antibacterial properties. Frontiers in microbiology. 2023; 14: 1135308. https://doi.org/10.3389/ fmicb.2023.1135308.
- Rattanata N, Klaynongsruang S, Leelayuwat C, Limpaiboon T, Lulitanond A, et al. Gallic acid conjugated with gold nanoparticles: antibacterial activity and mechanism of action on foodborne pathogens. International journal of nanomedicine. 2016; 11: 3347-3356. https://doi.org/10.2147/IJN.S109795.
- Nguyen T L A, Bhattacharya D. Antimicrobial Activity of Quercetin: An Approach to Its Mechanistic Principle. Molecules (Basel, Switzerland). 2022; 27(8): 2494. https://doi.org/10.3390/molecules27082494.

- Zahara K, Bibi Y, Arshad M, Kaukab G, Al Ayoubi S, et al. In-vitro examination and isolation of antidiarrheal compounds using five bacterial strains from invasive species Bidens bipinnata L. Saudi Journal of Biological Sciences. 2022; 29(1): 472-479. https://doi. org/10.1016/j.sjbs.2021.09.006.
- 42. Yang QH, Yang J, Liu GZ, Wang L, Zhu TC, et al. Study on in vitro anti-tumor activity of Bidens bipinnata L. extract. African Journal of Traditional, Complementary and Alternative Medicines. 2013; 10(3). https://doi.org/10.4314/ajtcam.v10i3.24.
- Zhu L H, Qin RY, Guo H M, Ding X L, Guan X L, et al. The inhibitory effect of Binens bipinnata L. extract on U14 tumour in mice. African Journal of Traditional, Complementary, and Alternative Medicines: AJTCAM. 2013; 10(4): 66-69. https://doi. org/10.4314/ajtcam.v10i4.11.
- 44. Huang G, Tang B, Tang K, Dong X, Deng J, et al. Isoquercitrin inhibits the progression of liver cancer in vivo and in vitro via the MAPK signalling pathway. Oncology Reports. 2014; 31(5): 2377-2384. https://doi.org/10.3892/or.2014.3099
- Atta A H, Mouneir S M. Evaluation of some medicinal plant extracts for antidiarrhoeal activity. Phytotherapy Research. 2005; 19(6): 481-485. https://doi.org/10.1002/ptr.1639.
- 46. Wang X Y, Chen G R, Pan C X, Deng Z Y, Ge J F, et al. Polyacetylenes from Bidens bipinnata L. and their biological activities. Phytochemistry Letters. 2014; 7(1): 198-201. https://doi. org/10.1016/j.phytol.2013.12.002.
- 47. Bo Y, Yuan L P, Zhang J J, Meng D Di, Jing H, et al. Total flavonoids of Bidens bipinnata L. a traditional Chinese medicine inhibits the production of inflammatory cytokines of vessel endothelial cells stimulated by sera from Henoch-Schönlein purpura patients. Journal of Pharmacy and Pharmacology. 2012; 64(6): 882-887. https://doi.org/10.1111/j.2042-7158.2012.01480.x.
- 48. Yang X, Bai Z F, Zhang D W, Zhang Y, Cui H, et al. Enrichment of flavonoid-rich extract from Bidens bipinnata L. by macroporous resin using response surface methodology, UHPLC-Q-TOF MS/MS-assisted characterization and comprehensive evaluation of its bioactivities by analytical hierarchy process. Biomedical chromatography: BMC. 2020; 34(11): e4933. https://doi. org/10.1002/bmc.4933.

- 49. Kwon J-W, Byun E, Lee Eung-Joo, Kim Y-C, Jeong G-S, et al. Antioxidative and hepatoprotective effect of compounds from the flowers of bidens Bipinnata L. Korean Journal of Pharmacognosy. 2009; 40: 345-350.
- 50. Yang X, Bai Z F, Zhang Y, Cui H, Zhou H L. Flavonoids-rich extract from Bidens bipinnata L. protects pancreatic β-cells against oxidative stress-induced apoptosis through intrinsic and extrinsic pathways. Journal of Ethnopharmacology. 2021; 275. https:// doi.org/10.1016/j.jep.2021.114097.
- 51. Yang X W, Huang M Z, Jin Y S, Sun L N, Song Y, et al. Phenolics from Bidens bipinnata and their amylase inhibitory properties. Fitoterapia. 2012; 83(7): 1169-1175. https://doi.org/10.1016/j. fitote.2012.07.005.
- 52. Wang Y Q, Li S J, Man Y H, Zhuang G. Serum metabonomics coupled with HPLC-LTQ/orbitrap MS and multivariate data analysis on the ameliorative effects of Bidens bipinnata L. in hyperlipidemic rats. Journal of ethnopharmacology. 2020; 262: 113196. https://doi.org/10.1016/j.jep.2020.113196.
- Shen A Z, Li X, Hu W, Chen FH. Total flavonoids of Bidens bipinnata L. ameliorate experimental adjuvant-induced arthritis through induction of synovial apoptosis. BMC complementary and alternative medicine. 2015; 15(1): 437. https://doi. org/10.1186/s12906-015-0962-3.
- 54. Yuan L P, Chen F H, Ling L, Dou P F, Bo H, et al. Protective effects of total flavonoids of Bidens pilosa L. (TFB) on animal liver injury and liver fibrosis. Journal of ethnopharmacology. 2008; 116(3): 539-546. https://doi.org/10.1016/j.jep.2008.01.010.
- 55. Xue-ying B AO, Yuan Q X H, Yu-xiang L I U, Jing XI E, Ji-bing Q U N. Europrotection of Total Flavones of Bidens Bipinnata L. on Brain Damage Following Experimental Intracerebral Hemorrhage in Rats. Chinese Journal of Pharmacovigilance. 2010; 7(12): 710. https://www.zgywjj.com/EN/Y2010/V7/I12/710.